South Dakota State University

Open PRAIRIE: Open Public Research Access Institutional Repository and Information Exchange

Electronic Theses and Dissertations

2018

A Comprehensive Analysis of Aromatic-Proton Mediated Hydrogen Bonds

Mona S. Alshamrani South Dakota State University

Follow this and additional works at: https://openprairie.sdstate.edu/etd

Part of the Biochemistry Commons, and the Chemistry Commons

Recommended Citation

Alshamrani, Mona S., "A Comprehensive Analysis of Aromatic-Proton Mediated Hydrogen Bonds" (2018). *Electronic Theses and Dissertations*. 2434. https://openprairie.sdstate.edu/etd/2434

This Thesis - Open Access is brought to you for free and open access by Open PRAIRIE: Open Public Research Access Institutional Repository and Information Exchange. It has been accepted for inclusion in Electronic Theses and Dissertations by an authorized administrator of Open PRAIRIE: Open Public Research Access Institutional Repository and Information Exchange. For more information, please contact michael.biondo@sdstate.edu.

A COMPREHENSIVE ANALYSIS OF AROMATIC-PROTON MEDIATED

HYDROGEN BONDS

BY

MONA ALSHAMRANI

A thesis submitted in partial fulfillment of the requirements for the

Master of Science

Major in Chemistry

South Dakota State University

2018

A COMPREHENSIVE ANALYSIS OF *AROMATIC*-PROTON MEDIATED HYDROGEN BONDS

This thesis is approved as a creditable and independent investigation by a candidate for Master of Science in Chemistry degree and is acceptable for meeting the thesis requirements for this degree. Acceptance of this does not imply that the conclusions reached by the candidate are necessarily the conclusions of the major department.

| Suvobrata Chakravarty, Ph.D. Thesis Advisor | Date |
|--|------|
| Douglas Raynie, Ph.D. Head, Department of Chemistry and Biochemistry | Date |
| Dean, Graduate School Date | Date |

This thesis is dedicated to my father without you I would never be where I stand today.

ACKNOWLEDGEMENTS

I would like to thank numerous people in my life who have supported me. First, I would like to thank my mother for her love, patience, and her prayers for me. Many thanks to my husband for his help and encouragement. I would like to thank my two brothers, who have been always proud of me. I also would like to thank all my sisters for all the prayers, encouragement, and support. I would like to thank my mother-in-law for her support. Many thanks to my adviser Dr. Suvobrata Chakravarty for his patience, management, and care to complete this project successfully. Finally, I would like to thank Saudi Arabia, for their academic support and financial support.

CONTENTS

| LIST OF FIGURES: | vii |
|--|----------|
| LIST OF TABLES: | X |
| ABSTRACT | xi |
| CHAPTER 1. Introduction& Overview | 1 |
| 1.1. Significance of CHO hydrogen bond in biological system. | 1 |
| 1.1.1 Role of hydrogen bonds in protein: | 1 |
| 1.1.2 The function of hydrogen bonds in protein-DNA, and RNA stability. | 3 |
| 1.1.3 The effectiveness of hydrogen bond and in medical side: | 4 |
| 1.2 Objective: | 6 |
| 1.2.1 Choice of the dataset, and HB Geometry | 7 |
| 1.2.2 Hydrogen Bond Energy. | 10 |
| 2. Chapter 2. A Comprehensive Analysis of Aromatic-proton Mediated Bonds | Hydrogen |
| Abstract: | 13 |
| 2.1 Introduction | 14 |
| 2.2 Materials and methods: | 16 |
| 2.2.1 Identification of hydrogen bond geometry: | 16 |
| 2.2.2 Calculation of hydrogen bond criteria: | 18 |
| 2.2.3 Distance from point to the plane. (Highest of acceptor atoms) | 18 |
| | 19 |
| 2.2.4. Energy Calculations | 19 |
| 2.3 Results and discussion. | 20 |
| 2.3.1 Calculation of hydrogen bond criteria: | 20 |
| 2.3.2 Analysis of CD1-HD1O hydrogen bond. | 25 |

| 2.3.3 Analysis of acceptor atoms in CD1-HD1O hydrogen bond | 29 |
|--|----|
| 2.3.4 Case study. | 33 |
| 2.3.5 Interaction Energy | 40 |
| 2.4. Conclusion. | 43 |
| CHAPTER 3 | 44 |
| 3. Conclusions and Future Work | 44 |
| 3.1 Conclusions: | 44 |
| 3.2. Future Work | 45 |
| References: | 47 |
| Appendix 1 : | 53 |
| A.1.1 Input file extract from (PDB file) | 53 |
| A.1.2 output file(hydrogen bond parameters) | 59 |

| Figure 1.1 model for the contact between anti-HIV-1 the antibody (4E10) and the |
|--|
| membrane-proximal external region (MPER) of gp415 |
| Figure 1. 2 Ring atoms of Trp, Tyr, and Phe participate in hydrogen bonds interaction7 |
| Figure 1.3 Distance and angular parameters used when defining CHO hydrogen |
| bonds |
| Figure 1.4 Regular anion-quadrupole interactions and their energies11 |
| Figure2.1 The frequency of ring-atom participating in anion-quadrupole, HB-anion |
| quadrupole interactions of the Trp15 |
| Figure 2.2 The ring-atoms participating in HB interactions of the Trp16 |
| Figure 2.3 The geometrics criteria of hydrogen bond identification, A: Acceptor, D: |
| Donor,Ø Angle DHA17 |
| Figure 2.4 Formula of Distance from point to plane19 |
| Figure 2.5 (A) Density plot to show the distributions of distance of NE1-HE1, CD1-HD1, |
| and CA-HA bonds, (B) Charge of the atoms ring and there protons21 |
| Figure 2.6 Density plot of distribution of DA |
| Figure 2.7 Distributions of DHA the angle of all expected HB in the ring of Trp24 |
| Figure 2.8 Total number of hydrogen bond In CD1 and other carbon atoms in the |
| ring |

| in the ring25 |
|---|
| Figure 2.10 Histogram of the percentages of all expected hydrogen bond in Trp-NE1, CD1, |
| and CA26 |
| Figure 2.11 Ring plot of percentages of total Hydrogen bond of CD1, CZ2, CZ3, CE3, and |
| CH227 |
| Figure 2.12 The 75% cutoff of DA, DHA values in Trp-NE1O bond, (A):Sp2 |
| hybridization,(B): Sp3hybridization |
| Figure 2.13 Histogram of hydrogen bond in CD1 and other carbon atoms after applying |
| NE1 75% cutoff criteria |
| Figure 2.14 (A) distribution of Acceptor atoms around NE1, (B) distribution of Acceptor |
| atoms around CD1 |
| Figure 2.15 Acceptor atoms side chain that are making hydrogen bond with CD132 |
| Figure 2.16 Acceptor atoms main chain that are making hydrogen bond with CD133 |
| Figure 2.17 Histogram of the percentages of expected hydrogen bonding in Tyr34 |
| Figure 2.18 Histogram of the percentages of all expected hydrogen bonding in Phe34 |
| Figure 2.19 The DA of NE1-O, CD1-O, CZ2-O, CZ3-O, CE3, and CH2 in 3700 high- |
| resolution dataset |
| Figure 2.20 Distribution of percentages of expected hydrogen bond in ring of Trp from |
| 3700 non-redundant dataset |

| Figure 2.21 Histogram of all expected HB of Trp-CA, Trp-NE1, Trp-CD1 from3700 |
|---|
| dataset |
| Figure 2.22 The percentages of all expected HB in CD1 and Other carbon atoms in |
| Tyr |
| Figure 2.23 The Percentages of all HB in CD1 and Other carbon Atoms in Phe40 |
| Figures 2.24 Ring atom participation frequency and interaction energy41 |
| Figure 2.25 The most frequent hydrogen bond acceptors side chain |
| Figure 3.1 Structure of an HIV45 |
| Figure 3. 1 Lipid Binding Site in 4E10 CDRH1 Region in Co-crystal Structures |

LIST OF TABLES:

| Table 1.1 CHO hydrogen bond parameters defined in the literature to identify C-HO |
|---|
| Interactions9 |
| Table 2.1 results of D-H in Trp20 |
| Table 2.2 All expected HB in Trp from 3700 non-redundant single chains dataset |
| Table 2.3 Condition of DA of NE1 in HBPLUS calculation which is between 2.3_3.5, this |
| condition have been applied to CD1 and other Carbon atoms in Trp37 |
| Table 2.4 Percentages of all expected HB of Trp-CA, Trp-NE1, Trp-CD1 from3700 non- |
| redundant single chains dataset |

ABSTRACT

A COMPREHENSIVE ANALYSIS OF AROMATIC-PROTON MEDIATED HYDROGEN BONDS

MONA ALSHAMRANI

2018

Hydrogen bonds play critical role in folding, structure and recognition of biological macromolecules (e.g., proteins, RNA, DNA). In addition to classical hydrogen bonds (e.g., -OH---O=, -OH---O-, -NH---O- etc.), structural analysis of protein and nucleic acids, almost a decade ago, showed that hydrogen bonds (e.g., CH---O) with hydrogen atoms on aliphatic carbon atoms (hereafter, *aliphatic*-protons) also play very important role in the structure and function of biomolecules. Even though, protons of aromatic ring systems (hereafter, *aromatic*-protons) are more polar than the *aliphatic*-protons, systematic analysis of hydrogen bonds of *aromatic*-protons have not been carried out. Therefore, I carried out a systematic analysis of hydrogen bonds that are made with *aromatic*-protons of tryptophan, tyrosine and phenylalanine amino acids in high-resolution structures of proteins and their complexes using a computer program that I created in R- and Perl languages. The analysis showed that aromatic CH---O are very common in proteins and tryptophan-CD1 is the most frequent participant in *aromatic* CH---O hydrogen bonds. The normalized frequency of occurrences of *aromatic* CH---O hydrogen bonds are greater than that of the *aliphatic* CH---O hydrogen bonds. Therefore, like *aliphatic* CH---O hydrogen bonds we anticipate that *aromatic* CH---O hydrogen bonds are likely to play equally, if not more, important role in macromolecular folding, stability and recognition. The numerous

examples of *aromatic* CH---O hydrogen bonds observed here provides a good source of information that can be probed further by experiments to validate their important contribution.

CHAPTER 1. Introduction& Overview

1.1. Significance of CH---O hydrogen bond in biological system.

Hydrogen bonds (H-bonds) happen in the hydrogen of a highly electronegative atoms such as nitrogen, oxygen, fluorine etc.[1].H-bonds are naturally weaker than most covalent bonds, but collectively they can make a huge contribution in the function and stability of biological macromolecules (e.g., all H-bonds of double stranded DNA holds the entire genome). Occasionally, carbon atoms have not been measured a conventional hydrogen bond donor because of carbon has low electronegativity compared to oxygen and nitrogen. In addition to electronegative atoms (e.g., N, O, Cl, F etc.), several studies over the past two decades have established that carbon atoms are also capable of forming weak hydrogen bonds (CH---O hydrogen bonds)[1, 2]. These studies emphasize the important role of CH---O bonds in protein stability and function such as activation, recognition and enzyme catalysis[2, 3].

1.1.1 Role of hydrogen bonds in protein:

In structural characterizations of proteins and protein–protein interfaces[4], analysis of hydrogen bonds is usually the first step of understanding how the protein might function. In general, similar analysis of CH---O H-bonds suggested that they could contribute to the protein stability and protein-protein recognition [5, 6]. There are many studies described the CH---O hydrogen bonding interaction at the protein-protein interface in the *aliphatic*-protons[7], and the calculations showed that the average energy contribution of H-bond is 30%; that of a CH---O hydrogen bonds is 17%; and that of a hydrophobic interactions 50%[7, 8]. Values of CH---O hydrogen bonds can reach as high as ~ 40–50%

in some complexes[7]. Also, this study have reported these results, CH---O hydrogen bonds repeatedly exist between adjacent strands in both parallel and antiparallel orientations, having the obvious structural motif of bifurcated H-bonds [5, 7]. Additionally, this study submits that the weak CH---O hydrogen bonds makes an important influence to the association and stability of protein complexes [3, 7-10]. Therefore, the weak CH---O hydrogen bonds makes an important contribution to the association and stability of protein complexes and protein-protein interaction [5, 7, 8]. In addition, In protein-protein binding hydrogen bonds often possibly show a powerful role and hence their designs can be used more directly to recognize native from nonnative binding orientations. Their patterns of interactions not only dictate specific recognition, but also provide much of the stability[3, 5]. Moreover, a recent study of USP7 catalytic domain for activation shows that CH---O hydrogen bond is one of the important factors that modulate USP7cd activation[11], comparative analysis of Inactive and active USP7 states shows that CZ2-HZ2---O in W285 makes hydrogen bond and this is strongly accepted that CH---O hydrogen bond is very important to activation of protein[11]. Furthermore, a comprehensive database analysis of CH---O hydrogen bonds in 3124 alpha-helices and their corresponding helix stations investigate the occurrence of CH---O hydrogen bonds in alpha-helices and helix termini in globular proteins. This analysis shows that CH---O hydrogen bonds contribute to the classical –NH---O hydrogen bonds and other interactions to the overall strength and stability of helix and therefore of proteins [3].

1.1.2 The function of hydrogen bonds in protein-DNA, and RNA stability.

More importantly, C-H---O hydrogen bonds play very effective roles in protein-DNA recognition, as well as DNA and RNA stability[12]. Indeed, CH---O hydrogen bonds contribute to the stability of Z-DNA[13]. The stability of A- and B-DNA comes from the stacking of the bases on top of each other. Otherwise, left-handed Z-DNA is stabilized by 04'---H6-C6 hydrogen bond[8, 14] [13]. A strong polarization of the guanine bases in Z-DNA is consistent with the Z-DNA specific guanine 06 and N7 coordination to metal and organic cations. The proximity of its N2 and C8 positions are neighboring phosphate groups, as well as several other Z-DNA-specific conformational features are there [13, 14]. Therefore, CH---O hydrogen bond weak interaction, but it still play significant roles in biological systems. For instance, in protein-DNA complexes, hydrogen bond play role in protein-DNA recognition[15]. A study shows that, on analysis of protein-DNA interfacial hydrogen bonds in 107 crystal structures using the in-house program "pyrHBfind" indicates that an overall smaller number of hydrogen bonds led to the consideration of alternative mechanisms playing a decisive role in stabilizing such complexes [16, 17]. These include CH---O hydrogen bonds [15]. Thus, in binding of proteins to DNA regulatory CH---O interactions play powerful role in the control of many cell processes [4, 13, 17, 18]. Group of researchers investigated and analyzed the CH---O interactions in the protein-DNA interface, based on 43 crystal structures of protein-DNA complexes[18]. They were surprising about the results, they have established that, the number of close intermolecular CH---O hydrogen bond contacts involving the thymine methyl group and position C5 of cytosine is equal to the number of protein-DNA hydrogen bonds involving nitrogen and oxygen atoms as donors and acceptors[18]. After they did a comprehensive

analysis they have reported that C5 of cytosine and C5-Met of thymine form fairly weak CH---O hydrogen bonds with Asp, Asn, Glu, Gln, Ser, and Thr, contributing to the specificity of recognition[18]. Therefore, the classical protein-DNA hydrogen bonds enable to enhance the stability of such complexes[19]. Likewise, there are several factors that can affect selectivity of nucleotide incorporation by a DNA polymerase, base pair hydrogen bonding, base pair size and shape, and base stacking interactions[15, 20]. However, all of these factors are necessary but not enough to achieve optimal efficiency for DNA polymerase comparing to hydrogen bond[12]. Therefore, CH---O hydrogen bond is weak bonds but in this case it is very important factor for efficiency and specificity of the human mitochondrial DNA polymerase[15, 18].

1.1.3 The effectiveness of hydrogen bond and in medical side:

Significantly, several studies have revealed that CH---O hydrogen bonds are responsible for determining of folded protein including enzymes and antibodies[4, 21]. As a result, CH---O bonds will be useful interaction in designing effective antibiotics, antibody, and antigen[21] [9]. In 2013 Lavanya and co-workers have studied the role of CH---O interactions in β -lactamases and the Influence of this bond on the structural stability[22]. In this study, they have studied the effect of CH---O interactions on major specificity and stability of β -lactamases in relation to environmental favorites. They have found that, all the residues located in the active site of β -lactamases are involved in CH----O interactions. These results used in understanding of the stability of β -lactamases[22]. In addition, CH---O hydrogen bonds act as an important driving force in ligand selectivity in the conformational stability of penicillin binding proteins[23], CH---O interactions contribute about 20-25% of total hydrogen bonds and this result will be useful for researcher to explore penicillin binding proteins[22]. In addition to all the above, recent studies have proved the significance, and effectiveness of CH---O hydrogen bonds in develop new antigens for the design of HIV vaccine in the near future. Wilson and colleagues show that 4E10 recognizes an extended epitope that Includes gp41 MPER and viral membrane lipids and identify specific lipid-binding regions[24]. In this study there are many factors led to identify the specific lipid-binding areas, however hydrogen bonds where the TRP residue HD1-atom is the hydrogen bond donor was one of these factors in this study[4, 21]. In Fig (1.1), they have seen that superposition of the 4E10 Fabs bound to 06:0 PA and to peptide epitope respectively, revealed that CDRH3 in its lipid-bound conformation would form unfavorable steric clashes with the gp41 peptide in the joining site[21, 24]. Although Lys100E (H) and Lys683 (P) could easily adopt different conformations, Trp680 (P) located in a restricted pocket, was within 1.3–1.7A° of Leu100C[24].



Figure1.1 model for the contact between anti-HIV-1 the antibody (4E10) and the membrane-proximal external region (MPER) of gp41[25].The model showing the unusually long CRDH3 with the two Trp residues (W100 and W110b) that engage the antigen was inspired from the recent study of Rujas [24], and was created using the antibody pdb-ids, 5x08 (in complex with the mutant antigen), 2fx7 (absence of lipids) and 4xcc (in presence of phosphatidylcholine). The proximity of HCDR3 Trp residues to the lipid head groups could be facilitated by anion–quadrupole interactions[25].

Supplementary, CH---O hydrogen bond plays significant role in developing a new antigens for the design of HIV vaccine[21]. In the study of Peptidomimetics built inhibitor design for HIV on of the results in this study examine the role of hydrogen bond which is the cutting results that show six hydrogen bonds formation between the ligand and binding interface of HIV-gp120. According to all of these studies [21, 23, 26], CH---O H-bonds are typically weaker than conventional hydrogen bonds, but this interaction has been considered very important. Hence, there is a critical requirement of studying this type of interaction and expose the common occurrence of CH---O weak hydrogen bonds in protein structures.

1.2 Objective:

Protons of aromatic ring systems (hereafter, *aromatic*-protons) are more polar than the *aliphatic*-protons, systematic analysis of hydrogen bonds of *aromatic*-protons have not been carried out. As a result, the objective of this project was to study and analysis the Aromatic-proton mediated hydrogen bonds, particularly Trp-CD1-HD---O and then we apply this study to other aromatic amino acids (Tyr, and Phe). Trp contains an α -amino group, an α -carboxylic acid group, and a side chain *indole*, making it a non-polar *aromatic* amino acid [6], also phenylalanine, tyrosine are all aromatic amino acids as the side chains are formed by aromatic rings. In order to accomplish this objective, we examine the criteria of hydrogen bond between the proton in Trp, Tyr, and Phe with acceptor atoms. In Fig (1.2) shows ring atom of W,Y, and F participate in hydrogen bonds interaction that we will consider in this project.



Figure 1. 2 Ring atoms of Trp, Tyr, and Phe participate in hydrogen bonds interaction.

1.2.1 Choice of the dataset, and HB Geometry.

A dataset involving 360 PDB files of very high resolution crystal structure of with resolution ≤ 1 Å, and a non-redundant set of 3700 high-resolution protein structures SCD dataset. We consider this data because with resolution values of 1 Å or so, are highly ordered and it is easy to see every atoms in the electron density map also we will be able to see the protons [6]. Lower resolution structures, with resolution of 3 Å or higher, show only the basic outlines of the protein chain, and the atomic structure must be inferred. Then, we extract aromatic amino acids from the data set because we were only looking for hydrogen bond in this group. Next step, we characterize CH---O hydrogen bonds in the ring atoms of aromatic amino acids by application of one of the most commonly used methods. It has been used for measuring CH---O parameters (angles and distances) [11, 27]. In many studies, almost all of hydrogen bond calculations have been done by using different software programs such as HBPLUS. However, this program has not been used in the study of CH---O Aromatic-proton hydrogen bonds. HPBLUS calculates hydrogen bonds of O, N atoms. In this project in order to accomplish this objective of studying the hydrogen bond's parameters, I have developed a new method of studying the parameters of CH---O hydrogen bonds in the ring of aromatic amino acids. Although, in order to identify hydrogen bonds we used the suggested geometric criteria of parameters that are accepted for CH---O hydrogen bonding by various people who have worked in this field[28], and that summarized in the Fig (1.3), Table 1.1



Figure 1.3 Distance and angular parameters used when defining CH---O hydrogen bonds.

Typical van der Waals distances d (2.7 Å) and D (3.7 Å) are frequently used as distance cutoffs for hydrogen bond identification. Example, the accepted definitions of distance and angle accepted for CH---O hydrogen bonding summarized in the table below[29].

Table 1.1 CH---O hydrogen bond parameters defined in the literature to identify C-H...O

 Interactions.

| Distance | Distance | Angle | Angle | Reference | |
|-----------------------------|----------------------------------|----------------------|-----------------|--------------------------|-----------------------|
| C _α O (Å) | HO(Å) | (C _a -HO) | (HO=C) | | |
| 3.4 ± 0.2 2.6 ± 0.2 | 26 ± 0.2 | 134 ± 22 | - | Chakrabarti et al | |
| | 2.0 ± 0.2 | | | (protein)1998 | |
| 3.15 | 3.15 2.63 to 2.79 100-109 91-110 | 91-110 | Berman et al | | |
| 5.15 | | 71-110 | (collagen)1996. | | |
| 3 /3 | 2 50 | >90 | _ | Steiner (small | |
| 5.45 | 15 2.30 290 | - | molecules) 1994 | | |
| ~2.5 | | - >130 - | | Fabiola et al (β sheets) | |
| <3.5 | - | | ~150 | - | 1997 |
| 3.0 to 4.0 | 0 to 4.0 2.2 to 2.5 >90 - | >00 | 2 2 to 2 5 >00 | | Wahl et al. (DNA/RNA) |
| 5.0 10 4.0 | | -90 - | 1997 | | |
| 3 0 to 1 0 <2 8 | >110 | 120 to 140 | Desiraju (small | | |
| 5.0 10 4.0 | | 120 10 140 | molecules)1996 | | |

The parameters which were calculated include the following:

- The length of bond between the C atom and the H atom. (D-H)
- The distances between the Carbon atom and the O atom (DA)
- The angle at H, the C-H...O angle (DAH)

Different geometric formation standards have been developed to predict hydrogen bonds occurrence in the protein molecules by [28].

1.2.2 Hydrogen Bond Energy.

Hydrogen bond energy depends on the chemistry of the donor and acceptor atoms as well as their location, based on the potential function[30, 31]. Many studies have considered the characteristics of hydrogen bond using ab initio methods. It is necessary to determine the energy of hydrogen bond because energy considered to be one of the important factors for deciding a good/strong hydrogen bond [30, 32, 33]. Most of quantum chemical calculations and computations were performed by the Gaussian program.

Capabilities of Gaussian (similar to other large- scale computational chemistry packages[31, 34, 35]:

- Determine most stable (optimum) molecular geometry and energy. (Used to calculate ΔE and Keq.)
- Define a potential energy surface by stepping through a range of values for a geometry coordinate, such as bond distance or torsion angle.
- Predict IR, Raman, UV, NMR, and other spectra.
- Optimize transition states.
- Solvate molecules using the "polarized continuum (PCM)" or other models.

- Special tools for optimizing transition metal complexes, and other molecules containing large atoms.
- Model surfaces using a 2D periodic boundary condition (PBC) method, or crystals using 3D PBC.

In addition, the energy scale has been developed to revision hydrogen bonds occurrence in the molecules, they have defended the energy for weak CH---O interactions (-1.0 kcal/mol) and strong (-10.0 kcal/mol) [31, 33]. In a comperhasive analysis of anion quderpol interaction in aromatic-proton we have calculated the energy of anion quderpole hydrogen bonds[36]. The distributions of the energies suggest that the strength of the interactions, on average, is similar for all of the aromatic residues and that the average energy of Trp interactions is greater (more negative) than that of the others. On the basis of the calculated energy, we loosely defined weak (at least -3.0 kcal/mol) and strong (at most -8.0 kcal/mol) interactions Fig (1.4). This calculation was a driving force to study CH---O *aromatic*-proton mediated hydrogen bonds.



Figure 1.4 Regular anion–quadrupole interactions and their energies. (A) Distance (\leq 4.5 Å) and angle (\leq 35°) detection criteria. (B) Distribution of the interaction energies for Phe (top), Tyr (middle),and Trp (bottom). Energy ranges for the strong and weak anion– quadrupole interactions are indicated by the gray background. (C)Comparison of the distribution of properties between strong and weak interactions: distances (top), angles (left, bottom), and interplanar angles (right, bottom)[25].

2. Chapter 2. A Comprehensive Analysis of Aromatic-proton Mediated Hydrogen Bonds

Abstract:

In the biological macromolecules, CH---O hydrogen bonds make a contribution to the protein stability and protein-protein recognition and illustrate protein-protein interfaces[3, 4]. The study and analysis of this interaction is significant, possibly because it has been less investigated since CH---O interactions have been considered weak interactions. A comprehensive statistical analysis of C-H---O hydrogen in a very highresolution 360 PDB with resolution ≤ 1 Å, and A non-redundant set of 3700 high-resolution protein structures dataset in order to investigate the *aromatic*-proton creating hydrogen bonds. In this study, the CH---O hydrogen bonded of tryptophan interactions with aromatic rings in proteins have been studied by calculating DH, DA, and DAH parameters see Fig (2.1). Systematic analysis of hydrogen bonds of *aromatic*-protons have been accomplished in this study using three different geometrical parameters: the C–H distance (DH), C–O distance (DA) and C-H-O angle (DAH). I studied CH---O interactions of the aromaticproton, and I have observed that CD1-HD1...O interactions show the highest propensity, with Trp, Tyr, and Phe rings. In this analysis, only those contacts with the angle $(90^\circ \le \emptyset \le$ 180°) were accepted. In addition, bonds associated within a distance 2.3 - 3.5 Å were established as a hydrogen bond. Furthermore, In a comprehasive analysis of an anion quderpol interaction in an aromatic-proton we have calculated the energy of anion quderpole hydrogen bonds[25, 36]. The distributions of the energies suggest that the strength of the interactions, on average, is similar for all of the aromatic residues and that the average energy of Trp interactions is greater (more negative) than that of the others. On

the basis of the calculated energy, we loosely defined weak (at least -3.0 kcal/mol) and strong (at most -8.0 kcal/mol) interactions. The energy calculation shows the polarity of the Trp-CD1 atom anions. Consequently, this consists of the CD1-HD1---O interaction geometry and the energy calculation of anion quderpol interaction showing the capability of Trp-CD1 to make a hydrogen bond after Trp- NE1 atom, and the polarity of HD1-atoms to be a H-bond donor in the *aromatic* ring.

2.1 Introduction.

CH---O is one of the non-covalent bonds and it is defined as a special type of weak hydrogen bond [1, 33, 37]. These interactions often happen in the structures of biomolecule like amino acid, protein, DNA and RNA. A number of reports indicate the significance of these type of hydrogen bonds in stabilizing nucleic acid, protein structures, and protein -DNA interface. Furthermore, the weak CH---O bond makes an influential role to the relationship and stability of protein complexes and protein-protein interaction [17, 30].On the medical side, CH---O interactions can be important in drug design; for example, this interactions contribution in the direction of the structural stability and specificity of therapeutic proteins [9, 35, 38].CH---O interactions provide strong proof of the impact of this interaction in protein modeling, ligand design, and structure-activity analysis as well as information on the relative strength of various aromatic CH donors [25, 39]. More significantly, the importance in the study of the role of anion-quadrupole interactions in aromatic rings of (Phe),(Tyr),(Trp),in molecular recognition, folding, and assembly [25]. We have looked how the role of anions hydrogen bonds of tryptophan and tyrosine side chains and Trp-CD1 shows highest percentages in the energy calculation. Also, in the

regular anion-quadrupole interactions the frequency of a ring-atom participating in an anion-quadrupole and HB-anion-quadrupole interactions of Trp-CD1 shows highest percentages, see Fig (2.1)[25].



Figure2.1 The frequency of ring-atom participating in anion–quadrupole and HB-anion– quadrupole interactions of the Trp[25].

However, we still need serious study of CH---O interaction in the rings of aromatic amino acids, particularly in the *aromatic*-protons where the HD1-atom is the hydrogen bond donor. Therefore, in this research I scientifically accomplished a systematic analysis of hydrogen bonds that are made with *aromatic*-protons of tryptophan, tyrosine and phenylalanine amino acids in high-resolution structures of proteins and their complexes using a computer program that I created in R- and Perl languages in protein and the ringatoms participating in HB interactions of the Trp that I consider in this study are showing in Fig (2.2).Thus, this study would also be beneficial in the study of other weak, noncovalent interactions in biological macromolecules and the program that I have developed will be useful in the study of CH---O geometry for al researchers.



Figure 2.2 The ring-atoms participating in HB interactions of the Trp.

2.2 Materials and methods:

2.2.1 Identification of hydrogen bond geometry:

We selected a very high-resolution 360 PDB with resolution ≤ 1 Å, and A nonredundant set of 3700 high-resolution protein structures SCD dataset in order to investigate the CH---O in *aromatic* rings. The principles used for the selection of the high-resolution 360 PDB data set were solved with resolution ≤ 1 Å because we need to see the protons to be able to investigate the standard H-bonding criteria for CD1-HD1...O Interactions and other carbon atoms in the *aromatic* ring which are C---H distance (D-H), C---O distance (DA) and C—H—O angle (DAH). Then I have calculated all expected HB in A nonredundant set of 3700 high-resolution protein structures SCD dataset by study and analysis the CD1-HD1...O of Trp, Tyr, and Phe to validate the method that I developed to study CD1-HD1...O geometry .Importantly, I developed a program in R and Perl languages to recognize the CH---O in the aromatics amino acids in the data set that I have described above by determine this H-bonding criteria, and These parameters are represented in Fig (2.2).



Figure 2.3 The geometrics criteria of hydrogen bond identification, A: Acceptor, D: Donor,

Ø Angle DHA.

To conclude, the geometrics criteria of hydrogen bond identification that I have followed were: D-A \leq 3.5 Å, D-H \leq 1.5 Å, and 90° $\leq \emptyset \leq$ 180°, and only the bonds that satisfy these conditions were accepted as hydrogen bond.

2.2.2 Calculation of hydrogen bond criteria:

Hydrogen donor protein atoms analyzed in this study were HD1 of Trp CD1, Tyr CD1, and Phe CD1; acceptor atoms were oxygen, and carboxyl oxygen of the main chain and side chain of amino acids in the protein structure. Mainly, I used the 3d coordinate X, Y and Z in the PDB files of CD1, and HD1 atoms and then I have created a script in R program to calculate the distance (DH) using the coordinate geometry formula of the distance. The distance between points (x_1, y_1, z_1) and (x_2, y_2, z_2) is:

$$d = \sqrt{(x_2 - x_1)^2 + (y - y_1)^2 + (z_2 - z_1)^2}$$

Where d is the distance.

Similarly, we calculate the distance (DA) of CD1, and acceptor atoms were *oxygen*, and *carboxyl oxygen* of the main chain and side chain. Furthermore, I have written script in R program to find the angle (DHA) between three atoms. I use the formula below:

$$\theta = \cos^{-1} \frac{\overrightarrow{a \ b}}{|a| \ |b|}$$

Where \xrightarrow{a} is the vector between hydrogen atom and the acceptor, \xrightarrow{b} is the vector between hydrogen atom and carbon atom, and θ is the angle between \xrightarrow{a} and \xrightarrow{b} .

2.2.3 Distance from point to the plane. (Highest of acceptor atoms).

Likewise, I have written a script in R to calculate the shortest distance in space between given point and a plane for NE1 and CD1 in TRP amino acid to see the distribution of acceptor atoms, and I use the formula in Fig (2.4) [29],to calculate the distance from point which are acceptor atoms to plane which are CD1,and NE1. Finally, I have looked at the distribution of these acceptors around each plane.

The distance D between a point $P_0(x_0, y_0, z_0)$ and the plane ax + by + cz + d = 0is $D = \frac{|ax_0 + by_0 + cz_0 + d|}{\sqrt{a^2 + b^2 + c^2}}$

Figure 2.4 Formula of Distance from point to plane

2.2.4. Energy Calculations

The energy of interactions was computed using NWChem quantum mechanical calculations[31]. For computing the energy of interaction (e.g., for an anion–quadrupole pair) the coordinates of side chain atoms of the interacting pair were first extracted from a *p*rotein *d*ata *b*ank (PDB) file. Hydrogen atoms for these side chains were added using the Open Babel version 2.3.1 chemical toolbox. Each CB atom had three protons and setting the pH to 7.0 (in Open Babel) ensured that Asp/Glu carboxyl groups remained deprotonated. An interacting pair of the type **A–B** (e.g., Asp–Phe) had three input files: (i) **A**, (ii) **B**, and (iii) **A–B**, where the **A** input file has the coordinates of the atoms of **A**. Each of the three input files for a pair was imported to the NWChem 6.6 program for energy calculations. Single-point energies were calculated using DFT with the Grimme DFT-D3 dispersion correction43 with the AUG-CC-PVDZ basis set and B3LYP functional[40]. The energy (in EH units) was extracted from the corresponding NWChem log files[33]. The

energy of interaction ($_EAB$) was obtained as, $_EAB = E(A-B) - [E(A) + E(B)]$, and $_EAB$ in EH units was then converted to kcal/mole. There were 6,256 HB-anion–quadrupole A– **B** pairs in total for the energy calculations[30] [25].

2.3 Results and discussion.

2.3.1 Calculation of hydrogen bond criteria:

Using a very high-resolution (\leq 1Å) 360 crystal structures PDB files. A simple process was developed for identifying CD1-HD1...O interactions in proteins. I run the script that I have written (see the method) to calculate the first geometric parameter of HB which is the distance between CD1.....HD1 (the distance (D-H)).I have calculated this distances for all ring atoms in Trp see Table (2.1).

Table2.1 results of D-H in Trp

| Atoms | Distance DH |
|---------|-------------|
| NE1-HE1 | 0.84 - 0.99 |
| CD1-HD1 | 0.92 - 0.99 |
| CZ2-HZ2 | 0.92 - 0.98 |
| CZ3-HZ3 | 0.92 - 0.99 |
| СЕЗ-НЕЗ | 0.92- 0.98 |
| СН2-НН2 | 0.91- 0.98 |
| СА-НА | 0.97 – 0.99 |



From *CHARMM36*, the charge of NE1 in the nature is (- 0.51) and the second higher charge is CD1 atom (-0.15) see Fig (2.4). I expect that, the length of CD1- HD1 will be similar or almost close to NE1-HE1. However, the result was disappointed CD1 atom shows longer distance than NE1 atom furthermore CD1 atom shows length close to the other carbon atoms, and the reason was because of the polarity and the charge of NE1 is more than CD1 .In addition, CA---HA shows the longer distance because CA the charge of alpha carbon is very weak comparing to NE1 and CD1 see Fig (2.4). The conclusion, *aromatic*-protons in the ring system are higher charge than the *aliphatic*-protons Fig (2.4).



Figure 2.5 (**A**) Density plot to show the distributions of distance of NE1-HE1, CD1-HD1, and CA-HA bonds, (**B**) Charge of the atoms ring and there protons.

After that, I used the program that I developed (see the method) to calculate and analysis the second geometric parameter of HB, the distance (DA) which is the distance between ring atoms and the acceptor CD1---O. Adding, all acceptor atoms were (e.g. –OH, –COO⁻, C=O) of the main chain and side chain that I considered hydrogen bonds only the acceptors with $2.3 \le DA \le 3.58$ Å Fig (2.5).



Figure 2.6 Density plot of distribution of DA of the atoms ring of Trp,CD1, CZ2,CZ3,CE3, and CH2.

In the analysis of 360 very high resolution protein structure, I have calculated the third geometric parameter of HB which is C—H—O angle (DAH) in Trp-CD1 and other carbon atoms in the ring. Only the acceptor atoms under this condition ($90^{\circ} \le \theta \le 180^{\circ}$) were accepted as hydrogen bonds acceptor atoms Fig (2.6).



Angle CE3-HE3...O
Figure 2.7 Distributions of DHA the angle of all expected HB in Trp-CD1, Trp-CZ2, Trp-CZ3, Trp-CH2 and Trp-CE3.

Successively, I make comparison in terms of total number of acceptor atoms for each atoms in the ring of Trp, CD1, CZ2, CZ3, CE3, and CH2. I have found the total number of hydrogen bond of acceptor atoms under the two conditions: $(2.3 \le DA \le 3.58$ Å) and $(90^\circ \le D-H-A \le 180^\circ)$ for CD1 more than other carbon atoms see Fig (2.8).



Figure 2.8 Total number of hydrogen bond In CD1, CZ2, CZ3, CE3, and CH2.

Then, I normalize the data and the percentage of total expected HB and in Trp-CD1 44% which is 20% more than Trp-CZ2 .Then I apply CD1 criteria to other carbon atoms in the ring just to make the result more accurate Fig (2.9).



Figure 2.9(**A**) Total number of all expected HB In Trp-CD1 comparing to Other carbon atoms in the ring with (DA 2.3 \leq DA \leq 3.58 Å), and (90° \leq D-H-A \leq 180°) : (**B**) Total number of all expected HB In Trp-CD1 comparing to Other carbon atoms with (DA 2.71 \leq DA \leq 3.58 Å.), and (90° \leq D-H-A \leq 179°) which is CD1 criteria.

2.3.2 Analysis of CD1-HD1...O hydrogen bond.

To provide more evidence to the previous results which is the capability of CD1 to make hydrogen bond, I have done some test based on the distance (DA) and the angle (DHA) of this bonds. First test, I compare the result of Trp-CD1 mediate hydrogen bond with a weak hydrogen bond (*alpha*- carbon) Trp-CA with a strong hydrogen bond Trp-NE1.I have selected alpha-carbon because many report have showed the ability of alpha

carbon (CA) to make hydrogen bond and that was surprising because the charge of alpha carbon in nature is very small (0.09) comparing to CD1(0.25) and NE1(0.5).Hence, I calculated DA and DAH of NE1, and CA atoms from the same PDB files that I used to study CD1. After that, I have calculated the percentages of all expected hydrogen bond of Trp-NE1, and Trp-CA and compare the result with hydrogen bond in Trp-CD1 Fig (2.10).



Figure 2.10 Histogram of the percentages of all expected hydrogen bond in Trp-NE1, CD1, and CA.

From the result of the first test above I have proofed the ability of *aromatic*- protons to form CH---O a hydrogen bond in particular Trp-CD1-HD1. Second test, analysis acceptor atoms which are oxygen, and carboxyl oxygen of the main chain and side chain of amino acids in the protein structure in CD1-HD1....O bonds. Conventionally, carbon has not been considered a predictable hydrogen bond donor due to its reasonably low

electronegativity compared with oxygen and nitrogen. In the meantime NE1 have been well studied, and there are some software available to calculate hydrogen bond in nitrogen. Therefore, I got the data of hydrogen bond of NE1 that have been calculated from HBPLUS and then I applied NE1 criteria to CD1 and other carbon atoms in the ring of Trp. Moreover, I applied the criteria of acceptor atoms with sp3, and sp2 hybridization in Trp-NE1....O hydrogen bond to create truthful comparison. In terms of accepter atoms with Sp3 hybridizations: DA of HB of NE1 atoms were 2.3-3.43 Å, and DAH the angle of HB of NE1 atoms were between 98.7°- 179.4°. Accepter atoms with Sp2 hybridizations: DA of HB of NE1 atoms were 2.42-3.43 Å, and DAH of HB of NE1 atoms were 98.7°-179.4°. In the two case sp3 and sp2 hybridizations. CD1 shows the highest percentage comparing to other carbon atoms in the Trp ring Fig (2.11).



Figure 2.11 Ring plot of percentages of total Hydrogen bond of CD1, CZ2, CZ3, CE3, and CH2 after applying NE1 criteria to them. A: accepter atoms with Sp3 hybridizations, B: Accepter Atoms with Sp2 hybridizations.

Second test, I have done cut off to see where the most population will be for each of DA, and DHA In Trp-NE1 bonds when Sp2, and Sp3 hybridization. I have found that from 75 % CUTOFF most of the bonds almost have DA between 2.82 - 3.09 Å and for the DHA 135 °- 170° in Sp2, in terms of Sp3 most of the bonds almost have DA between 2.80 - 3.06Å, and DHA between 130° -175° Fig (2.12). Besides, I use quantile function in R Program to get the exact 75% cutoff values.



B



Figure 2.12 The 75% cutoff of DA, DHA values in Trp-NE1....O bond, (A): Sp2 hybridization,(B): Sp3 hybridization.

After that, I use the criteria of 75% cutoff of DA, DHA of Trp-NE1....O and I apply it to DA and DHA of Trp-CD1, Trp-CZ2, Trp-CZ3, Trp-CE3, and Trp-CH2. as a result, CD1 shows the highest percentages comparing to other carbon atoms in the ring Fig (2.13).



Figure 2.13 Histogram of total hydrogen bond in CD1 and other carbon atoms in the ring after applying NE1 75% cutoff criteria.

2.3.3 Analysis of acceptor atoms in CD1-HD1...O hydrogen bond.

In the calculation of the shortest distance in space between given point (acceptor), and plane which are Trp-NE1, and Trp-CD1, the highest of all acceptor to the two plan:Trp-NE1, and Trp-CD1 were from 0.002 to 2. Before I take the absolute values as the formula require see Fig (2.14) in the method, I use the result and make plot to look at the distributions of acceptor atoms around the plan. Also, I have decided a vector in the plan and see the distribution of the acceptor of CD1 and NE1 around this vector because CD1

atom close to NE1 atom in the ring ,and NE1 has high electronegativity I want to identify if the polarity of CD1 because of this feature. The distribution and the number of acceptor atoms around each plane (NE1, CD1) were equal. As a result, the polarity of CD1 is not because of the position of this atom in the ring Fig (2.14).





Figure 2.14 (A) distribution of Acceptor atoms around NE1, (B) distribution of Acceptor atoms around CD1, The number of acceptor atoms are equal around each plane.

In the analysis of acceptor atoms that I have found in hydrogen bond calculation of CD-HD1...O bond, I analyze all side chain acceptor atoms that are making hydrogen bond with CD1 Fig (2.15) and main chain Fig (2.16). CD1 show the highest percentage in all acceptor type in main chain, however, in side chain CD1 was the highest in all of them but when the acceptor is Gln, Trp-CZ2 make hydrogen bonding with this type of acceptor more than CD1. In general, in both case side chain and main chain CD1 show higher number in making hydrogen bonding.

Acceptor (Side Chain).

| Atoms | ASP | GLU | ASN | <mark>GLN</mark> | SER | THR | TYR |
|-------|-----|-----|-----|------------------|-----|-----|-----|
| CD1 | 57 | 33 | 31 | <mark>11</mark> | 37 | 25 | 17 |
| CZ2 | 25 | 31 | 9 | <mark>19</mark> | 21 | 16 | 21 |
| CZ3 | 6 | 15 | 5 | 8 | 13 | 9 | 16 |
| CH2 | 13 | 28 | 5 | 11 | 11 | 12 | 10 |
| CE3 | 14 | 11 | 17 | 7 | 13 | 4 | 11 |



Figure 2.15 acceptor atoms side chain that are making hydrogen bond with CD1.



Figure 2.16 acceptor atoms main chain that are making hydrogen bond with CD1.

2.3.4 Case study.

First, In order to examine CD-HD1...O bonding, I have repeated all the study and the calculation I have done to analysis Trp-CD1 in 360 PDB files to Tyr-CD1, and Phe-CD1. After I run the program and calculated all expected hydrogen bond in Tyr, total number of hydrogen bonding in Tyr-CD1 52%, and Tyr-CE1 40% Fig (2.17).



Figure 2.17 Histogram of the percentages of all expected hydrogen bonding I Tyr in the high resolution 360 PDB files.

In addition, in the study of hydrogen bonding in the atoms ring of Phe, total number of hydrogen bond in Phe-CD1 45%, Phe-CE1 27%, and Phe-CZ 26% see Fig (2.18).



Figure 2.18 Histogram of the percentages of all expected hydrogen bonding Phe in the high resolution 360 PDB files.

Second, I selected a non-redundant set of 3700 high-resolution protein structures SCD dataset in order to study the CD1-HD1...O interactions of Trp rings. I calculated all expected hydrogen bond Trp-CD1-HD1...O in this data set using the same method to study the distance between CD1---O which is (DA) and the only acceptors satisfy the condition of regular hydrogen bond DA \leq 3.5Å accepted as hydrogen bond Fig (2.19).

TRP-NE1...O



TRP-CD1...O











TRP-CH2...O



Figure 2.19 The DA of NE1-O, CD1-O, CZ2-O, CZ3-O, CE3, and CH2 in 3700 high-resolution protein structures SCD dataset.

As well, I have calculated all expected HB in NE1 and other carbon atoms in the ring to make comparison with CD1, CD1 make more hydrogen bond after NE1 and this support our previous results Table (2.2). Moreover, HBPLUS calculate DA of NE1 between **2.3-3.5**Å.I apply this condition to CD1 and other Carbon atoms in Trp and after apply this condition CD1 still show the higher number of hydrogen bond after NE1 Table (2.3), Fig (2.20).

| Atoms | Total HB DA≤3.5 | Percentages |
|---------|-------------------|------------------|
| TRP-NE1 | <mark>6000</mark> | <mark>59%</mark> |
| TRP-CD1 | 5442 | <mark>53%</mark> |
| TRP-CZ2 | 2212 | 22% |
| TRP-CZ3 | 1585 | 15% |
| TRP-CH2 | 1616 | 16% |
| TRP-CE3 | 1870 | 18% |

Table 2.2 All expected HB in Trp from 3700 non-redundant single chains dataset.

Table 2.3 Condition of DA of NE1 in HBPLUS calculation which is \leq **3.5** Å.I apply this condition to CD1 and other Carbon atoms in Trp.

| Atoms | Percentages |
|---------|------------------|
| TRP-NE1 | <mark>56%</mark> |
| TRP-CD1 | <mark>39%</mark> |
| TRP-CZ2 | 14% |
| TRP-CZ3 | 9% |
| TRP-CH2 | 10% |
| TRP-CE3 | 11% |



Figure 2.20 Distribution of percentages of expected hydrogen bond in ring of Trp from 3700 non-redundant single chains dataset.

In addition, alpha carbon considered as weak atom, and it can make hydrogen bond as many reports have been done in the past. Because of that, I have discovered all estimated HB in CA from3700 non-redundant single chains dataset, and then I compare the result with Trp-NE1 and Trp-CD1. CD1 was capable to make hydrogen bond after NE1 and more than CA. Therefore,CD1 second high charge and second high electronegativity atom in the ring comparing to other atoms in the ring of Trp Table(2.4), Fig(2.21).

 Table 2.4 Percentages of all expected HB of Trp-CA, Trp-NE1, Trp-CD1 from3700 non

 redundant single chains dataset



Figure 2.21Histogram of all expected HB of Trp-CA, Trp-NE1, Trp-CD1 from3700 nonredundant single chains dataset.

Finally, In the analysis and study of hydrogen bonding geometry, I have repeated all the calculation I have done to analysis Trp-CD1 in the data 3700 non-redundant single chains dataset to look at Tyr-CD1, and Phe-CD1. After I run the program and calculated all expected hydrogen bond in Tyr, total number of hydrogen bonding in Tyr-CD1 23%, and Tyr-CE1 11% Fig (2.22).



Figure 2.22 The percentages of all expected HB in CD1 and Other carbon atoms in Tyr.

Furthermore, I have calculated all expected HB in Phe from 3700 non-redundant single chains dataset. I have seen that Trp-CD1 has highest percentages in forming hydrogen bond, and that give more evidence to our results see Fig (2.23).



Figure 2.23 The Percentages of all HB in CD1 and Other carbon Atoms in Phe.

2.3.5 Interaction Energy.

In the study of anion–quadrupole interactions of the tyrosine (Tyr) and tryptophan (Trp) rings [25].We examined whether the computed energies are consistent with the frequency with which interactions occur in protein crystal structures[30, 33, 40]. For example, the FATOM_ds values for tyrosine, phenylalanine, and tryptophan atoms are consistent with the computed energy(e.g. FATOM_ds for Phe-CZ is 18% while that for Phe-CD1 is 22%) see Figures (2.24 A, B, C)[25].



Figures 2.24 Ring atom participation frequency and interaction energy.(A) The height of the bar (FATOM_ds) adjacent to the name of each atom represents the frequency (%) of its participation in anion– quadrupole interactions for that residue type, Trp (top), Tyr (bottom left), and Phe (bottom right). The number within the bracket above or below each bar represents the interaction energy (kilocalories per mole) when the indicated atom is closest to the interacting anion. (B) The distribution represents the energy of anion–quadrupole interactions when Trp-CD1 (top) and Trp-CH2 (bottom) atoms are closest to the interacting anion. The dotted lines with an arrow represent the mean interaction energy in kilocalories per mole. (C) Linear correlation (r = 0.82) between the energy and FATOM_ds for tryptophan ring carbon atoms. For a qualitative agreement between the energy and the frequency, we use FATOM_ds in place of log(FATOM_ds)[25]

In addition, the energy of an anion-quadrupole interaction based on quantum mechanical calculations depends not only on the interaction geometry but also on the ring atom[31, 37], the frequency of aromatic ring participation in anion-quadrupole. In The aspartate/glutamate anion- quadrupole interactions, Asp/Glu side chain carboxylates were the most frequent hydrogen bond acceptors for each type of side chain hydrogen bond donor in proteins [25]. Since the energy calculation takes time to finish, I have showed the statistical analysis of the frequent hydrogen bond acceptors for each type of side chain, main chain hydrogen bond donor in proteins in the 3700 dataset that I have been used to study the bond geometry. In the statistical analysis, I have recognized that: first, most frequent hydrogen bond side chain and main chain showing in CD1 atom for Trp, Tyr, and Phe. Second, the aspartate/glutamate CD1 hydrogen bond acceptors see Fig (2.23). Therefore, I have estimated that the energy calculation should consist with this statistical analysis as shown Fig (2.25).



Figure 2.25 The most frequent hydrogen bond acceptors side chain

2.4. Conclusion.

The high-resolution set of 360 protein with a resolution ≤ 1 Å, and a non-redundant set of 3700 high-resolution protein structures SCD dataset were statistically analyzed by focusing mainly on the study of *aromatic*-proton mediated hydrogen bonds in terms of geometry; distance, and angle of the bond also, the statistical analysis of the frequent hydrogen bond, acceptors and the energy calculation of the hydrogen bond in anion quadrupole interaction. Therefore, the comprehensive analysis of *aromatic*-proton have been completed in this study illustrate the capability of Trp-CD1 to make a hydrogen bond after a Trp- NE1 atom. Additionally, this analysis validate the polarity of a HD1-atoms to be H-bond donor in CD1-HD1....O interaction.

Acknowledgments

The authors would like to acknowledge the South Dakota State University academic computing facility for the access to the Linux cluster for all the energy calculations. The authors would like to thank Suvobrata Chakravarty, Brian Moore, Jay Shore, and Adron R. Ung for all the energy calculation have been use in this study.

CHAPTER 3

3. Conclusions and Future Work

3.1 Conclusions:

In this study, the method was a program in R and Perl to study the geometries of CH---O hydrogen bonds in aromatic amino acids in a very high-resolution 360 protein structure with resolution ≤ 1 Å, and A non-redundant set of 3700 high-resolution protein structures SCD dataset. This method successfully finds all expected hydrogen bond contacts within distance DA ≤ 3.5 Å, and angle DHA $\leq 90^{\circ}$. This method with high simplicity and efficiency studied CH---O hydrogen bonds in *aromatic*-protons in a very big dataset. Additionally, the method reasonably give results that display the ability of a CD1 atom in the ring of tryptophan to make hydrogen bonds, were other available software to do that have difficulties in accomplishing that for two main reasons: (1) the data of protein that we use is very large, (2) many programs find the parameter of bonds in high electronegative atoms such as oxygen and nitrogen. However, the script still has small drawbacks that need to be overcome, such as timing the code takes 2 to 3 hours to finish the calculation when datasets are very large, and R software works very well with txt file or csv files. Therefore, we need to resave the PDB file in the appropriate file. In general, the program shows the capability to calculate all expected hydrogen bonds and I can just make a small change in the function that I used to speed up or convert the script that I use in this program to another language programs such as Perl, or C^{++} .

3.2. Future Work

I scientifically studied CH---O hydrogen bonding in Trp, Tyr, and Phe in a highresolution, and non-redundant protein single chains, and I clearly recognize the capability of CD-HD1 in the rings of aromatic amino acids to make hydrogen bond. In addition, we have observed the usefulness of Trp-CD1-HD1 in designing effective antibiotics and antibodies. For example, in HIV structure[41], see Fig (3.1)[42], one study reported that the 4E10 was able to bind with the gp41 in HIV, see Fig(3.2)[24].Also, in the study of Peptidomimetics built inhibitor design for HIV, one of the results in this study examine the role of hydrogen bonds which have critical results that show six hydrogen bonds forming between the ligand and binding interface of HIV-gp120 [26].Therefore, my upcoming goal is to look at the interfaces protein–protein, DNA/RNA–protein, and membrane–protein.



Figure 3.1 Structure of an HIV[42].



Figure 3. 2 Lipid Binding Site in 4E10 CDRH1 Region in Co-crystal Structures [24, 25].

In addition, building a webserver is very important since there is a need for programs to calculate the parameter of CH---O hydrogen bond in a large datasets. Then, the webserver will help many researchers to analyze the CH---O hydrogen bonds in ether aromatic or aliphatic protons.

References:

- Chopra, P.P.a.D., Understanding of Noncovalent Interactions Involving Organic Fluorine. 2015: p. 37-61.
- Seiji Tsuzuki, K.H., Tadafumi Uchimaru, Masuhiro Mikami, and and K. Tanabe, *The Magnitude of the CH/π Interaction between Benzene and Some*

Model Hydrocarbons. ACS, 2000. 122: p. 3746-3753.

3. Sethumadhavan*, V.S.a.R., *Effect of C-H...O Interactions in the Structural*

Stability of "All-Alpha" Proteins. ChemTech, 2011. Vol.3: p. PP 348-356.

- Daniel Boamah, T.L., Franchesca A. Poppinga, Shraddha Basu,[†] Shahariar Rahman,[†] and a.S.C. Francisca Essel, *Characteristics of a PHD Finger Subtype*. ACS, 2018. 57: p. 525–539.
- C. Nick Pace, 2* Hailong Fu,1 Katrina Lee Fryar,2 John Landua,2, et al., Contribution of hydrogen bonds to protein

stability. the protein society March 2014.

- Alok Jain, V.R., 2, a.R. Sankararamakrishnan1, and *, Lone pair … π interactions between water oxygens and aromatic residues: Quantum chemical studies based on high-resolution protein structures and model compounds. PROTEIN SCIENCE, 2009. 18: p. 595-605.
- Lai, L.J.a.L., *CH---O Hydrogen Bonds at Protein-Protein Interfaces*. JBC 2002.
 277(4): p. 37732–37740.
- Lai[‡], L.J.a.L., *CH....O Hydrogen Bonds at Protein-Protein Interfaces*.
 BIOLOGICAL CHEMISTRY, 2002. Vol. 277(40): p. 37732–37740.

- 9. SANDRO MECOZZI, A.P.W., JR., AND DENNIS A. DOUGHERTY*, *Cation-1 interactions in aromatics of biological and medicinal interest: Electrostatic potential surfaces as a useful qualitative guide.* PNAS. Vol. 93: p. 10566-10571.
- 10. Thomas Steiner, G.K., *Hydrogen bonds with* π *-acceptors in proteins: frequencies and role in stabilizing local 3D structures1*. 2001. **305**(3): p. 535-557.
- Ays, eg€ul O[°] zen, 2 Lionel Rouge[′], 3 Charlene Bashore, 1 Brian R. Hearn, 4
 Nicholas J. Skelton, 2,* and Erin C. Dueber 1, 5,*, Selectively Modulating
 Conformational States of
- *USP7 Catalytic Domain for Activation*. cell press, January 2018. **Volume 26**(issue 1): p. p72–84.e7, 2.
- Hisae Tateishi-Karimata1, Tatsuya Ohyama1,[†], Takahiro Muraoka2, Peter Podbevsek3,, S.T. Adam M. Wawro2, Shu-ichi Nakano5, Kazushi Kinbara2, Janez Plavec3, and and Naoki Sugimoto1, *Newly characterized interaction stabilizes DNA structure: oligoethylene glycols stabilize G-quadruplexes CH-π interactions*. Nucleic Acids Res, 2017. **45**(12): p. 7021–7030.
- 13. Rich, A.H.a.A., *The Biology of Left-handed*
- *Z-DNA*. JBC, 1996. **271**(17): p. 11595–11598.
- GESSNER§, M.E.A.R.V., Stereoelectronic effects of deoxyribose 04' on DNA conformation. Proc. Natl. Acad. Sci. USA January 1995. Vol. 92: p. 180-184.
- Shayantani Mukherjee, S.M., † and Dhananjay Bhattacharyya*, *Role of Hydrogen* Bonds in Protein-DNA Recognition: Effect of Nonplanar Amino Groups. J. Phys, March 17, 2005. 109: p. 10484-10492.

 Yael Mandel-Gutfreund1, H.M., Robert L. Jernigan2 and a.V.B. Zhurkin2*, A Role for CHO Interactions in

Protein-DNA Recognition. JMB, 1998. 277: p. 1129-1140.

17. Yael Mandel-Gutfreund1, H.M., Robert L. Jernigan2 and a.V.B. Zhurkin2*, A Role for CH.....O Interactions in

Protein-DNA Recognition. JMB, 1998. 277: p. 1129-1140.

- BZhurkin2, Y.-G.H.R.L.V., A role for CH...O interactions in protein-DNA recognition. Journal of Molecular Biology, 1998 Volume 277(5): p. Pages 1129-1140.
- Modan K Das 1, a.H.-K.D., *A survey of DNA motif finding algorithms*. BMC Bioinformatics 2007.
- 20. Xing Du 1, Yi Li 1,[†], Yuan-Ling Xia 1, et al., *Insights into Protein–Ligand Interactions: Mechanisms, Models, and Methods.* Int. J. Mol. Sci, 2016. **17**(2).
- Tomonaga Ozawa *, E.T., Motoyasu Ozawa, Chiaki Handa, Harunobu Mukaiyama, and S.K. Toshihiro Nishimura, Kosuke Okazaki, *The importance of CH/p hydrogen bonds in rational drug design: An ab*

initio fragment molecular orbital study to leukocyte-specific protein

tyrosine (LCK) kinase. 2008. 16: p. 10311–10318.

- Anbarasu, P.L.S.R.A., Influence of C-H...O interactions on the structural stability of β-lactamases. JBP, 25 June 2013. 39: p. 649–663.
- Hang Yin1, a.A.D.F., 3, *Drugging Membrane Protein Interactions*. PMC, 2016
 Feb 5.

24. Adriana Irimia, A.S., Robyn L. and I.A.W. Stanfield, *Crystallographic Identification of Lipid as an Integral*

Component of the Epitope of HIV Broadly

Neutralizing Antibody 4E10. cell press, 2016 January. 44: p. 21–31.

- 25. Suvobrata Chakravarty, A.R.U., Brian G Moore, Jay S. Shore, and Mona Alshamrani, A Comprehensive Analysis of Anion–Quadrupole Interactions in Protein Structures. acs.biochem, 2018.
- Umashankar Vetrivel1*, P.S. and G.S. Naveen kumar Nagarajan2,
 Peptidomimetics Based Inhibitor Design

for HIV – 1 gp120 Attachment Protein. JPB, November 2009. Vol.2.

- 27. Shahbazi*, Z., *Mechanical Model of Hydrogen Bonds in Protein Molecules*.
 American Journal of Mechanical Engineering, 2015. 3: p. 47-54.
- Shahbazi*, Z., Mechanical Model of Hydrogen Bonds in Protein Molecules.
 American Journal of Mechanical Engineering, 2015, Vol.3: p. 47-54.
- 29. Rusczyk, R., Introduction to Geometry. 2007. 1.
- 30. Mutasem Omar Sinnokrot[†], a.C.D.S., *High-Accuracy Quantum Mechanical Studies of π-π Interactions in Benzene Dimers*. ACS, 2006. **110**: p. 10656–10668.
- 31. Andrei V. Afonin, a.A.V.V.a.M.V.S.b., *Estimating the energy of intramolecular hydrogen bonds from 1H NMR and OTAIM calculations*. 2016(Issue 47).
- Mondal A1, D.S., *Quantum mechanical electronic structure calculation reveals* orientation dependence of hydrogen bond energy in proteins. Proteins, 2017. 85: p. 1046–1055.

- 33. Kwang S. Kim*, P.T., and Jin Yong Lee, Molecular Clusters of π-Systems: Theoretical Studies of Structures, Spectra, and Origin of Interaction Energies. ACS, 2000. 100: p. 4145-4148.
- 34. Keller, J., Introduction to Gaussian. 2008: p. 1-37.
- Chenzhong Liao1, Markus Sitzmann1, Angelo Pugliese1, and Marc C Nicklaus1, Software and resources for computational medicinal chemistry. NIH, 2011. 8: p. 1057–1085.
- Houk, S.E.W.a.K.N., Are Anion/π Interactions Actually a Case of Simple Charge-Dipole Interactions? ACS, 2010. 114: p. 8658–8664.
- 37. Sławomir J. Grabowski, †,‡ W. Andrzej Sokalski,§ Edyta Dyguda,§ and Jerzy Leszczyn´ski, *Quantitative Classification of Covalent and Noncovalent H-Bonds*. ACS, 2006. 110: p. 6444-6446.
- 38. Sciences, I.J.o.P.a.P., *CH...O INTERACTIONS STABILIZE THE STRUCTURE OF THE THERAPEUTIC PROTEINS: A*
- *COMPUTATIONAL STUDY*. International Journal of Pharmacy and Pharmaceutical Sciences, 2011. **3**(3): p. ISSN- 0975-1491.
- Pierce AC1, S.K., Bemis GW., *Kinase inhibitors and the case for CH...O hydrogen bonds in protein-ligand binding*. Proteins: Structure, Function, and Bioinformatics, 2002. 49(4): p. 567–576.
- 40. Jr, J.H.A.D.M., *CHARMM36 all-atom additive protein force field: validation based on comparison to NMR data.* Journal of Computational Chemistry, 2013.
 34(25): p. 2135-2145.

- 41. Yunji W. Davenport, A.P.W., Jr., Pamela J. Bjorkman, *Structure of an HIV-2* gp120 in Complex with CD4. Journal of Virology, 2016. **90**.
- 42. Andrea Rubbert, G.B.a.M.O., Pathogenesis of HIV-1 Infection. 2011.

Appendix 1 :

A.1.1 Input file extract from (PDB file)

Atom

| Residue | name | Amino | Х | У | Z | ATOM | Filename |
|---------|------|-------|--------|--------|--------|------|-----------|
| 1 | Ν | VAL | -5.066 | 0.058 | 13.305 | Ν | 1AHOA.pdb |
| 1 | 0 | VAL | -3.354 | 2.058 | 13.22 | 0 | 1AHOA.pdb |
| 2 | Ν | LYS | -2.935 | 1.92 | 11.013 | Ν | 1AHOA.pdb |
| 2 | 0 | LYS | -2.14 | 3.055 | 8.683 | 0 | 1AHOA.pdb |
| 2 | NZ | LYS | 3.015 | -0.908 | 9.879 | Ν | 1AHOA.pdb |
| 3 | Ν | ASP | -0.952 | 4.628 | 9.7 | Ν | 1AHOA.pdb |
| 3 | 0 | ASP | 1.633 | 5.085 | 9.122 | 0 | 1AHOA.pdb |
| 3 | OD1 | ASP | -3.28 | 6.409 | 8.885 | 0 | 1AHOA.pdb |
| 3 | OD2 | ASP | -2.501 | 8.311 | 9.661 | 0 | 1AHOA.pdb |
| 4 | Ν | GLY | 1.236 | 5.367 | 6.913 | Ν | 1AHOA.pdb |
| 4 | 0 | GLY | 2.152 | 4.86 | 4.32 | 0 | 1AHOA.pdb |
| 5 | Ν | TYR | 4.317 | 5.073 | 4.948 | Ν | 1AHOA.pdb |
| 5 | 0 | TYR | 5.649 | 2.562 | 4.175 | 0 | 1AHOA.pdb |
| 5 | ОН | TYR | 6.666 | 10.964 | 3.42 | 0 | 1AHOA.pdb |
| 5 | HD1 | TYR | 5.81 | 7.066 | 1.498 | Н | 1AHOA.pdb |
| 6 | Ν | ILE | 4.167 | 2.716 | 2.493 | Ν | 1AHOA.pdb |
| 6 | 0 | ILE | 6.11 | 1.571 | 0.867 | 0 | 1AHOA.pdb |
| 6 | HD11 | ILE | 2.075 | 0.281 | -1.143 | Н | 1AHOA.pdb |

| 6 | HD12 | ILE | 0.885 | 1.098 | -0.401 | Н | 1AHOA.pdb |
|----|------|-----|--------|--------|--------|---|-----------|
| 6 | HD13 | ILE | 1.693 | 1.807 | -1.637 | Н | 1AHOA.pdb |
| 7 | Ν | VAL | 5.886 | -0.382 | 1.947 | Ν | 1AHOA.pdb |
| 7 | 0 | VAL | 5.968 | -3.045 | 1.368 | 0 | 1AHOA.pdb |
| 8 | Ν | ASP | 7.852 | -2.885 | 0.152 | Ν | 1AHOA.pdb |
| 8 | 0 | ASP | 8.789 | -4.536 | 2.05 | 0 | 1AHOA.pdb |
| 8 | OD1 | ASP | 10.659 | -3.821 | -0.483 | 0 | 1AHOA.pdb |
| 8 | OD2 | ASP | 10.724 | -4.16 | -2.677 | 0 | 1AHOA.pdb |
| 9 | Ν | ASP | 8.899 | -6.313 | 0.69 | Ν | 1AHOA.pdb |
| 9 | 0 | ASP | 11.365 | -7.556 | 2.999 | 0 | 1AHOA.pdb |
| 9 | OD1A | ASP | 10.052 | -8.373 | -1.046 | 0 | 1AHOA.pdb |
| 9 | OD1B | ASP | 7.259 | -9.001 | -0.022 | 0 | 1AHOA.pdb |
| 10 | Ν | VAL | 11.539 | -5.98 | 1.374 | Ν | 1AHOA.pdb |
| 10 | 0 | VAL | 14.076 | -3.57 | 1.917 | 0 | 1AHOA.pdb |
| 11 | Ν | ASN | 11.936 | -3.517 | 2.483 | Ν | 1AHOA.pdb |
| 11 | 0 | ASN | 12.527 | 0.005 | 2.272 | 0 | 1AHOA.pdb |
| 11 | OD1 | ASN | 11.592 | -0.56 | 5.466 | 0 | 1AHOA.pdb |
| 11 | ND2 | ASN | 13.78 | -0.071 | 5.346 | Ν | 1AHOA.pdb |
| 12 | N A | CYS | 11.777 | -1.274 | 0.601 | Ν | 1AHOA.pdb |
| 12 | N B | CYS | 11.503 | -1.503 | 0.812 | Ν | 1AHOA.pdb |
| 12 | 0 A | CYS | 9.398 | -0.462 | -0.917 | 0 | 1AHOA.pdb |
| 12 | O B | CYS | 9.28 | -0.529 | -0.459 | 0 | 1AHOA.pdb |
| 13 | Ν | THR | 10.311 | 1.539 | -0.709 | Ν | 1AHOA.pdb |
| | | | | | | | |

| 13 | 0 | THR | 9.341 | 1.771 | -3.313 | 0 | 1AHOA.pdb |
|----|-----|-----|--------|--------|---------|---|-----------|
| 13 | OG1 | THR | 10.462 | 4.274 | -1.095 | 0 | 1AHOA.pdb |
| 14 | Ν | TYR | 7.335 | 2.541 | -2.624 | Ν | 1AHOA.pdb |
| 14 | 0 | TYR | 6.256 | 4.851 | -4.232 | 0 | 1AHOA.pdb |
| 14 | ОН | TYR | 3.108 | -2.87 | -2.523 | 0 | 1AHOA.pdb |
| 14 | HD1 | TYR | 5.689 | 0.639 | -1.651 | Н | 1AHOA.pdb |
| 15 | Ν | PHE | 7.615 | 3.844 | -5.714 | Ν | 1AHOA.pdb |
| 15 | 0 | PHE | 5.604 | 4.519 | -7.468 | 0 | 1AHOA.pdb |
| 15 | HD1 | PHE | 10.063 | 7.004 | -7.486 | Н | 1AHOA.pdb |
| 16 | Ν | CYS | 6.165 | 6.679 | -7.332 | Ν | 1AHOA.pdb |
| 16 | 0 | CYS | 6.005 | 9.231 | -8.253 | 0 | 1AHOA.pdb |
| 17 | Ν | GLY | 4.113 | 8.823 | -9.417 | Ν | 1AHOA.pdb |
| 17 | 0 | GLY | 2.387 | 11.926 | -9.6 | 0 | 1AHOA.pdb |
| 18 | Ν | ARG | 1.558 | 9.847 | -9.618 | Ν | 1AHOA.pdb |
| 18 | 0 | ARG | -0.307 | 8.714 | -7.772 | 0 | 1AHOA.pdb |
| 18 | NE | ARG | -1.042 | 8.024 | -13.088 | Ν | 1AHOA.pdb |
| 18 | NH1 | ARG | 1.237 | 7.847 | -13.407 | Ν | 1AHOA.pdb |
| 18 | NH2 | ARG | -0.022 | 5.96 | -13.116 | Ν | 1AHOA.pdb |
| 19 | Ν | ASN | -1.363 | 10.688 | -7.674 | Ν | 1AHOA.pdb |
| 19 | 0 | ASN | -3.066 | 8.242 | -5.771 | 0 | 1AHOA.pdb |
| 19 | OD1 | ASN | -0.84 | 12.35 | -4.997 | 0 | 1AHOA.pdb |
| 19 | ND2 | ASN | -2.493 | 13.807 | -5.344 | Ν | 1AHOA.pdb |
| 20 | Ν | ALA | -3.65 | 9.042 | -7.798 | Ν | 1AHOA.pdb |

| 20 | 0 | ALA | -4.528 | 5.608 | -7.577 | 0 | 1AHOA.pdb |
|----|------|-----|--------|--------|---------|---|-----------|
| 21 | Ν | TYR | -2.707 | 6.534 | -8.541 | Ν | 1AHOA.pdb |
| 21 | 0 | TYR | -1.952 | 3.665 | -6.735 | 0 | 1AHOA.pdb |
| 21 | ОН | TYR | 2.947 | 1.179 | -8.287 | 0 | 1AHOA.pdb |
| 21 | HD1 | TYR | -0.564 | 2.998 | -10.246 | Н | 1AHOA.pdb |
| 22 | Ν | CYS | -1.3 | 5.749 | -6.206 | Ν | 1AHOA.pdb |
| 22 | 0 | CYS | -2.425 | 4.122 | -3.294 | 0 | 1AHOA.pdb |
| 23 | Ν | ASN | -3.474 | 5.741 | -4.437 | Ν | 1AHOA.pdb |
| 23 | 0 | ASN | -5.504 | 3.201 | -3.219 | 0 | 1AHOA.pdb |
| 23 | OD1 | ASN | -6.971 | 6.627 | -2.18 | 0 | 1AHOA.pdb |
| 23 | ND2 | ASN | -8.149 | 5.68 | -3.809 | Ν | 1AHOA.pdb |
| 24 | Ν | GLU | -4.881 | 3.516 | -5.341 | Ν | 1AHOA.pdb |
| 24 | 0 | GLU | -4.678 | 0.155 | -4.494 | 0 | 1AHOA.pdb |
| 24 | OE1A | GLU | -4.535 | 1.98 | -10.096 | 0 | 1AHOA.pdb |
| 24 | OE1B | GLU | -5.56 | 1.17 | -9.874 | 0 | 1AHOA.pdb |
| 24 | OE2A | GLU | -5.987 | 3.569 | -10.363 | 0 | 1AHOA.pdb |
| 24 | OE2B | GLU | -5.825 | -0.966 | -9.404 | 0 | 1AHOA.pdb |
| 25 | Ν | GLU | -2.903 | 1.441 | -4.966 | Ν | 1AHOA.pdb |
| 25 | 0 | GLU | -2.198 | -0.652 | -2.205 | 0 | 1AHOA.pdb |
| 25 | OE1 | GLU | 0.121 | -1.596 | -5.871 | 0 | 1AHOA.pdb |
| 25 | OE2 | GLU | -0.773 | -0.812 | -7.683 | 0 | 1AHOA.pdb |
| 26 | Ν | CYS | -2.686 | 1.527 | -2.178 | Ν | 1AHOA.pdb |
| 26 | 0 | CYS | -4.387 | -0.002 | 0.502 | 0 | 1AHOA.pdb |

| 27 | Ν | THR | -5.286 | 0.822 | -1.381 | Ν | 1AHOA.pdb |
|----|------|-----|---------|--------|--------|---|-----------|
| 27 | 0 | THR | -7.026 | -2.268 | -0.904 | 0 | 1AHOA.pdb |
| 27 | OG1 | THR | -7.464 | 0.694 | -3.28 | 0 | 1AHOA.pdb |
| 28 | Ν | LYS | -5.405 | -1.773 | -2.375 | Ν | 1AHOA.pdb |
| 28 | 0 | LYS | -4.919 | -5.054 | -1.109 | 0 | 1AHOA.pdb |
| 28 | NZ | LYS | -2.174 | -3.122 | -8.245 | Ν | 1AHOA.pdb |
| 29 | Ν | LEU | -3.945 | -3.106 | -0.474 | Ν | 1AHOA.pdb |
| 29 | 0 | LEU | -4.122 | -3.584 | 3.124 | 0 | 1AHOA.pdb |
| 29 | HD11 | LEU | -0.091 | -1.441 | 0.995 | Н | 1AHOA.pdb |
| 29 | HD12 | LEU | 0.92 | -2.715 | 0.884 | Н | 1AHOA.pdb |
| 29 | HD13 | LEU | 0.543 | -1.865 | -0.455 | Н | 1AHOA.pdb |
| 30 | Ν | LYS | -5.687 | -2.978 | 1.623 | Ν | 1AHOA.pdb |
| 30 | 0 | LYS | -7.351 | -1.4 | 4.412 | 0 | 1AHOA.pdb |
| 31 | Ν | GLY | -5.875 | -0.579 | 2.961 | Ν | 1AHOA.pdb |
| 31 | 0 | GLY | -7.834 | 1.093 | 2.161 | 0 | 1AHOA.pdb |
| 32 | Ν | GLU | -7.359 | 2.624 | 3.705 | Ν | 1AHOA.pdb |
| 32 | 0 | GLU | -9.139 | 4.301 | 1.198 | 0 | 1AHOA.pdb |
| 32 | OE1 | GLU | -8.544 | 7.631 | 4.503 | 0 | 1AHOA.pdb |
| 32 | OE2 | GLU | -10.213 | 7.269 | 5.813 | 0 | 1AHOA.pdb |
| 33 | Ν | SER | -7.085 | 4.864 | 1.908 | Ν | 1AHOA.pdb |
| 33 | 0 | SER | -4.61 | 5.66 | 1.789 | 0 | 1AHOA.pdb |
| 33 | OG | SER | -7.135 | 7.787 | 2.147 | 0 | 1AHOA.pdb |
| 34 | Ν | GLY | -4.831 | 6.815 | -0.123 | Ν | 1AHOA.pdb |

| 34 | 0 | GLY | -4.036 | 8.514 | -2.049 | 0 | 1AHOA.pdb |
|----|-----|-----|--------|--------|--------|---|-----------|
| 35 | Ν | TYR | -1.902 | 8.672 | -1.268 | Ν | 1AHOA.pdb |
| 35 | 0 | TYR | 0.677 | 8.786 | -1.814 | 0 | 1AHOA.pdb |
| 35 | ОН | TYR | 1.521 | 12.888 | 2.359 | 0 | 1AHOA.pdb |
| 35 | HD1 | TYR | -1.956 | 10.61 | 0.829 | Н | 1AHOA.pdb |
| 36 | Ν | CYS | 0.459 | 10.184 | -3.56 | Ν | 1AHOA.pdb |
| 36 | 0 | CYS | 2.291 | 12.516 | -3.602 | 0 | 1AHOA.pdb |
| 37 | Ν | GLN | 3.423 | 11.07 | -2.271 | Ν | 1AHOA.pdb |
| 37 | 0 | GLN | 6.363 | 11.613 | -2.559 | 0 | 1AHOA.pdb |

| Residue | Amino | ATOM | Ac residue | Ac amino | Ac ATOM | DH | DAH | DA | file |
|---------|-------|------|------------|----------|---------|----------|----------|----------|-----------|
| 318 | TRP | CD1 | 322 | GLU | OE1A | 0.922665 | 128.0838 | 2.713077 | 4N8GA.pdb |
| 318 | TRP | CD1 | 322 | GLU | OE2B | 0.923985 | 107.1814 | 2.772068 | 4N8GA.pdb |
| 311 | TRP | CD1 | 307 | THR | 0 | 0.925476 | 117.3123 | 2.965026 | 4YEPA.pdb |
| 234 | TRP | CD1 | 237 | ASP | 0 | 0.925804 | 138.9424 | 2.990718 | 4IILA.pdb |
| 257 | TRP | CD1 | 255 | VAL | 0 | 0.925864 | 96.87643 | 2.996869 | 4HNOA.pdb |
| 32 | TRP | CD1 | 31 | SER | OG B | 0.926417 | 108.0759 | 2.999742 | 3I94A.pdb |
| 669 | TRP | CD1 | 716 | SER | ОВ | 0.926901 | 120.4121 | 3.035548 | 4QLPB.pdb |
| 7 | TRP | CD1 | 168 | VAL | ОВ | 0.926983 | 139.1768 | 3.036643 | 2XU3A.pdb |
| 155 | TRP | CD1 | 151 | LEU | 0 | 0.927043 | 103.4604 | 3.039667 | 4PSRA.pdb |
| 348 | TRP | CD1 | 344 | ASN | 0 | 0.927089 | 110.574 | 3.041127 | 4QRNA.pdb |
| 314 | TRP | CD1 | 310 | ALA | 0 | 0.927101 | 98.6395 | 3.05029 | 4UABA.pdb |
| 157 | TRP | CD1 | 153 | ALA | 0 | 0.927389 | 102.864 | 3.055033 | 4FK9A.pdb |
| 153 | TRP | CD1 | 328 | TYR | OH | 0.927496 | 119.0355 | 3.061346 | 4JN7A.pdb |
| 269 | TRP | CD1 | 265 | ARG | 0 A | 0.927572 | 107.9995 | 3.068336 | 1MUWA.pdb |
| 94 | TRP | CD1 | 90 | ASP | 0 | 0.927648 | 133.9276 | 3.070831 | 4JN7A.pdb |
| 84 | TRP | CD1 | 76 | LEU | 0 | 0.927677 | 121.3262 | 3.075234 | 4HNOA.pdb |
| 149 | TRP | CD1 | 102 | ALA | 0 | 0.927744 | 120.7251 | 3.078645 | 5C5GA.pdb |
| 266 | TRP | CD1 | 261 | PRO | 0 | 0.927766 | 115.59 | 3.078825 | 5EJ8H.pdb |
| 81 | TRP | CD1 | 106 | ALA | 0 | 0.927809 | 157.3049 | 3.083514 | 4C08A.pdb |
| 150 | TRP | CD1 | 157 | GLY | 0 | 0.927911 | 112.2491 | 3.084567 | 4Z0YB.pdb |
| 238 | TRP | CD1 | 233 | GLN | 0 | 0.927916 | 97.90527 | 3.090767 | 5FLYB.pdb |
| 305 | TRP | CD1 | 307 | ASN | OD1 | 0.927935 | 124.2581 | 3.093184 | 4QP5A.pdb |
| 669 | TRP | CD1 | 716 | SER | O A | 0.928119 | 119.6893 | 3.098021 | 4QLPB.pdb |
| 109 | TRP | CD1 | 108 | ASP | OD2A | 0.928175 | 122.8109 | 3.113903 | 1MJ5A.pdb |
| 251 | TRP | CD1 | 250 | THR | OG1 | 0.928265 | 145.1888 | 3.114068 | 5JDAA.pdb |
| 119 | TRP | CD1 | 115 | ASP | 0 | 0.928291 | 140.6662 | 3.119354 | 4KEMA.pdb |
| 108 | TRP | CD1 | 56 | LEU | 0 | 0.928382 | 101.6619 | 3.121414 | 2VB1A.pdb |
| 159 | TRP | CD1 | 157 | SER | OG B | 0.928476 | 153.601 | 3.125962 | 4C08A.pdb |

A.1.2 output file(hydrogen bond parameters)
| 340 | TRP | CD1 | 337 | GLN | 0 | 0.928484 | 119.1491 | 3.126779 | 4WFOA.pdb |
|-----|-----|-----|-----|-----|-----|----------|----------|----------|-----------|
| 402 | TRP | CD1 | 398 | TRP | 0 | 0.92857 | 150.7976 | 3.127858 | 4G1QB.pdb |
| 165 | TRP | CD1 | 164 | LEU | 0 | 0.928632 | 134.4854 | 3.12955 | 5INJA.pdb |
| 156 | TRP | CD1 | 152 | TYR | 0 | 0.92864 | 126.3306 | 3.133462 | 4PP4A.pdb |
| 406 | TRP | CD1 | 408 | ALA | 0 | 0.928664 | 107.6677 | 3.135051 | 4G1QB.pdb |
| 194 | TRP | CD1 | 190 | THR | 0 | 0.9287 | 126.4022 | 3.135355 | 4XQCA.pdb |
| 343 | TRP | CD1 | 362 | THR | OG1 | 0.9287 | 113.4852 | 3.137228 | 4U9HL.pdb |
| 218 | TRP | CD1 | 216 | ASP | 0 | 0.928777 | 137.8681 | 3.137605 | 4ZBOA.pdb |
| 261 | TRP | CD1 | 294 | THR | 0 | 0.928813 | 137.6067 | 3.139696 | 5E9NA.pdb |
| 97 | TRP | CD1 | 96 | LEU | 0 | 0.92882 | 105.5705 | 3.143592 | 1W7CA.pdb |
| 235 | TRP | CD1 | 232 | ARG | 0 | 0.928821 | 91.21711 | 3.143779 | 4BJIA.pdb |
| 118 | TRP | CD1 | 112 | LEU | 0 | 0.928914 | 112.0337 | 3.143984 | 3WA2X.pdb |
| 88 | TRP | CD1 | 85 | LYS | 0 | 0.928971 | 138.9742 | 3.145527 | 4Z3GA.pdb |
| 222 | TRP | CD1 | 220 | TYR | 0 | 0.92898 | 98.72851 | 3.149459 | 4QOSA.pdb |
| 292 | TRP | CD1 | 288 | GLU | 0 | 0.928984 | 118.3828 | 3.151632 | 3U65B.pdb |
| 284 | TRP | CD1 | 282 | THR | OG1 | 0.928989 | 123.2822 | 3.153396 | 2FVYA.pdb |
| 574 | TRP | CD1 | 570 | SER | 0 | 0.929013 | 101.2095 | 3.154491 | 4C6FA.pdb |
| 153 | TRP | CD1 | 81 | ASN | OD1 | 0.929027 | 133.9615 | 3.155272 | 4G1QB.pdb |
| 32 | TRP | CD1 | 168 | TYR | 0 | 0.929047 | 154.7447 | 3.160221 | 4EYZA.pdb |
| 299 | TRP | CD1 | 295 | THR | 0 | 0.929052 | 153.0318 | 3.160936 | 4JCCA.pdb |
| 570 | TRP | CD1 | 219 | ASP | OD2 | 0.929069 | 102.6756 | 3.163705 | 4UDXX.pdb |
| 59 | TRP | CD1 | 55 | TYR | 0 | 0.929097 | 114.3212 | 3.164244 | 5COFA.pdb |
| 223 | TRP | CD1 | 181 | LEU | 0 | 0.929097 | 127.7496 | 3.164362 | 5HWQA.pdb |
| 223 | TRP | CD1 | 181 | LEU | 0 | 0.929098 | 128.048 | 3.165723 | 5HWOA.pdb |
| 891 | TRP | CD1 | 886 | ARG | O B | 0.929115 | 135.0887 | 3.167322 | 5C30A.pdb |
| 284 | TRP | CD1 | 282 | ASP | 0 | 0.92912 | 124.9162 | 3.168328 | 5DN9A.pdb |
| 72 | TRP | CD1 | 71 | PHE | 0 | 0.929122 | 143.1288 | 3.168702 | 4QQSA.pdb |
| 169 | TRP | CD1 | 165 | ARG | 0 | 0.929125 | 137.3148 | 3.170214 | 5CTMA.pdb |
| 149 | TRP | CD1 | 145 | THR | Ο | 0.929147 | 98.36793 | 3.172853 | 5C5GA.pdb |
| 71 | TRP | CD1 | 69 | ASP | Ο | 0.929147 | 103.0402 | 3.173134 | 2AXWA.pdb |
| 94 | TRP | CD1 | 90 | ASP | OD1 | 0.929151 | 125.886 | 3.174327 | 4JN7A.pdb |

| 69 | TRP | CD1 | 105 | THR | OG1 | 0.929151 | 108.6059 | 3.175796 | 5CL8A.pdb |
|-----|-----|-----|-----|-----|-----|----------|----------|----------|-----------|
| 233 | TRP | CD1 | 228 | SER | 0 | 0.929156 | 118.4408 | 3.176611 | 5A99A.pdb |
| 120 | TRP | CD1 | 117 | GLU | 0 | 0.929156 | 161.4718 | 3.176729 | 4CDJA.pdb |
| 133 | TRP | CD1 | 129 | GLU | 0 | 0.929157 | 154.8317 | 3.177471 | 4PSSA.pdb |
| 147 | TRP | CD1 | 179 | SER | OG | 0.929158 | 113.6795 | 3.179276 | 2YN0A.pdb |
| 227 | TRP | CD1 | 223 | ASN | 0 | 0.929159 | 140.3251 | 3.183185 | 4JN7A.pdb |
| 455 | TRP | CD1 | 451 | ASN | 0 | 0.929174 | 91.73961 | 3.18376 | 1LLFA.pdb |
| 305 | TRP | CD1 | 87 | SER | OG | 0.929183 | 108.1094 | 3.184883 | 4FK9A.pdb |
| 318 | TRP | CD1 | 314 | ARG | 0 | 0.929186 | 135.5485 | 3.186379 | 2JLIA.pdb |
| 308 | TRP | CD1 | 304 | THR | 0 | 0.929187 | 155.4781 | 3.18752 | 4MCOA.pdb |
| 337 | TRP | CD1 | 335 | ARG | 0 | 0.929201 | 104.9628 | 3.189624 | 1JFBA.pdb |
| 45 | TRP | CD1 | 43 | GLN | 0 | 0.929201 | 112.6999 | 3.189906 | 4REKA.pdb |
| 239 | TRP | CD1 | 235 | ASN | 0 | 0.929202 | 154.9397 | 3.189961 | 5FC1A.pdb |
| 389 | TRP | CD1 | 325 | THR | OG1 | 0.929208 | 108.5691 | 3.191436 | 4BQYA.pdb |
| 683 | TRP | CD1 | 696 | SER | 0 | 0.929213 | 163.3947 | 3.193117 | 4N5UA.pdb |
| 118 | TRP | CD1 | 72 | TRP | 0 | 0.929214 | 109.6235 | 3.193426 | 4L9OA.pdb |
| 455 | TRP | CD1 | 451 | ASN | OD1 | 0.929218 | 104.8101 | 3.194255 | 1LLFA.pdb |
| 28 | TRP | CD1 | 24 | SER | 0 A | 0.929219 | 123.9703 | 3.194823 | 2VB1A.pdb |
| 79 | TRP | CD1 | 75 | ARG | 0 | 0.929234 | 155.6359 | 3.195251 | 4PJ2C.pdb |
| 206 | TRP | CD1 | 207 | PRO | 0 | 0.929235 | 141.0161 | 3.196062 | 4MNCA.pdb |
| 203 | TRP | CD1 | 124 | ASP | OD2 | 0.929235 | 128.9655 | 3.196912 | 1Y93A.pdb |
| 59 | TRP | CD1 | 54 | LEU | 0 | 0.929239 | 108.2549 | 3.198957 | 5E75A.pdb |
| 243 | TRP | CD1 | 240 | ASN | 0 | 0.929241 | 137.5971 | 3.198983 | 3ZOJA.pdb |
| 250 | TRP | CD1 | 252 | ILE | 0 | 0.929252 | 117.2502 | 3.19957 | 1IXHA.pdb |
| 71 | TRP | CD1 | 68 | SER | 0 | 0.929258 | 139.5513 | 3.200212 | 4D0QA.pdb |
| 148 | TRP | CD1 | 146 | ARG | 0 | 0.929261 | 99.80256 | 3.204978 | 2NLRA.pdb |
| 215 | TRP | CD1 | 214 | SER | OG | 0.929263 | 168.5827 | 3.20589 | 5JBBS.pdb |
| 27 | TRP | CD1 | 23 | GLN | 0 | 0.929263 | 124.0881 | 3.207185 | 1TT8A.pdb |
| 212 | TRP | CD1 | 196 | GLY | 0 | 0.929265 | 119.0956 | 3.208561 | 4XDUA.pdb |
| 76 | TRP | CD1 | 74 | ASN | 0 | 0.929267 | 114.7568 | 3.20933 | 4UE8A.pdb |
| 215 | TRP | CD1 | 214 | SER | OG | 0.929272 | 167.5711 | 3.209393 | 5JB9S.pdb |

| 344 | TRP | CD1 | 355 | ASN | 0 | 0.929272 | 110.2114 | 3.212218 | 4X9YA.pdb |
|-----|-----|-----|-----|-----|-----|----------|----------|----------|-----------|
| 529 | TRP | CD1 | 316 | THR | 0 | 0.929272 | 109.509 | 3.213004 | 4D7CA.pdb |
| 119 | TRP | CD1 | 115 | ASP | OD1 | 0.929273 | 132.1677 | 3.213836 | 4KEMA.pdb |
| 327 | TRP | CD1 | 313 | LEU | 0 | 0.929274 | 118.4685 | 3.214053 | 4FK9A.pdb |
| 17 | TRP | CD1 | 13 | PRO | 0 | 0.929278 | 152.6694 | 3.214665 | 3NZLA.pdb |
| 196 | TRP | CD1 | 181 | GLN | 0 | 0.929278 | 118.7033 | 3.215138 | 4MTMA.pdb |
| 294 | TRP | CD1 | 306 | SER | OG | 0.929281 | 115.7075 | 3.215153 | 4X33B.pdb |
| 192 | TRP | CD1 | 150 | ASP | 0 | 0.929286 | 146.8763 | 3.21578 | 4YAPA.pdb |
| 303 | TRP | CD1 | 300 | ASN | 0 | 0.929287 | 111.0369 | 3.218121 | 1W7CA.pdb |